548 1-13 Gould

LIFE HISTORY AND ECOLOGY
OF
DAPHNIA PULEX SSP. PULICOIDES
WOLTERECK 1932

by

BLAINE W. LE SUER

A THESIS

Submitted to the Graduate Faculty

in

partial fulfillment of the requirements

for the degree of

Master of Science in Botany

at

Montana State College

Approved:
Head, Major Department
Chairman, Examining Committee
Dean, Graduate Division

Bozeman, Montana August, 1959

TABLE OF CONTENTS

LIST ACKNO	OF TA	LLUSTRA ABLES GMENTS	• •			•	•	•	•	•	•	•	•	•	•		•	•	•	•	•	•	•	•	•	•	•	111 iv
PART	I. :	INTROD	JCTI	ON		•	•				•			•		•	•			•	•	•	•					1
PART	Life	METHODS Histo:	ry S	tud	y				٠							•		•	•			•	٠	٠	•	٠	٠	2
	Graz: Resp:	ing St iration Oxyg e n	n St	udi	86	٠		٠	٠	٠		٠	٠	•	٠	٠	٠	٠	•	٠	٠	•	٠	٠	•	٠	٠	2 3 4 4
		Carbo	n Co n Di	oxio	npı de	Pr)11 (OC	luc	cti	or	ר ניינו)et	tei	rmi	ina	• iti	or	· ns	•	•	•	:	•	•	:	•	•	
PART	III. Life	RESU Histo	ry S	tud	У	٠	٠		•	•			٠	٠	٠	•	•	•	•	٠	٠	٠	•	•	•	*	•	7
		Insta: Growtl Durat:	n pe	er II	nst	tar	:		•	٠	•	•	•		٠	•	•	•	•	*	٠	•	٠	•	•		•	77 77 77
		Repro Morta	duct lity	ion Ra	te			•	•	•	•		•	•	•	•		•	•	•	•	•	•	•	•	•	•	11 13
	Graz:	ing St iratio	udy n St	udi	• e s	:	•	•	*	•	•	•	•	•	•	•	•	•	•	•	•		•	•	•		•	15 19
PART	Life Graz	DISCUS Histo ing St iratio	ry S udy	tud	У.	•		•	•	•	•	•	•	•	•	•	•	•	•	•	•	•	•	•	•	•	•	27 27 28 29
PART	٧.	SUMMAR	Υ.			•	•		٠	•	٠		•	•		•	٠			•	٠	•	•	•	٠		•	30
ITTE	RATUR	E CITE	D.								٠																	32

LIST OF ILLUSTRATIONS

<u>Figure</u>		Pag	<u>e</u>
1	Instar-weight Relationship	. 8	
2	Average Increase in Length of Each Instar	. 9	
3	Average Time in Days Required for Passage from One Instar to the Next	. 10	
4	Average Number of Eggs Formed and Average Number of Young Released per Reproductive Instar	. 12	
5	Survivors per Instar	. 14	
6	Average Filtering Rates of <u>Daphnia pulex</u> at Each Temperature Level		
7	Average Rate of Oxygen Consumption and Carbon Dioxide Production by Instar	. 23	
8	Average Rate of Oxygen Consumption and Carbon Dioxide Production on a per Animal Basis at Each Temperature Level	. 24	
9	Average Rate of Oxygen Consumption and Carbon Dioxide Production per Unit Weight at Each Temperature Level	. 25	

LIST OF TABLES

Number		•	Page
I	DATA ON INSTARS ONE THROUGH TWENTY-SIX	•	15
II	VOLUME OF WATER FILTERED BY <u>DAPHNIA</u> INSTARS ONE THROUGH TEN AT FIVE DEGREES CENTIGRADE	•	16
III	VOLUME OF WATER FILTERED BY <u>DAPHNIA</u> INSTARS ONE THROUGH TEN AT TEN DEGREES CENTIGRADE	•	16
IV	VOLUME OF WATER FILTERED BY <u>DAPHNIA</u> INSTARS ONE THROUGH TEN AT FIFTEEN DEGREES CENTIGRADE	•	17
V	VOLUME OF WATER FILTERED BY <u>DAPHNIA</u> INSTARS ONE THROUGH TEN AT TWENTY DEGREES CENTIGRADE	•	17
VI	VOLUME OF WATER FILTERED BY <u>DAPHNIA</u> INSTARS ONE THROUGH TEN AT TWENTY-FIVE DEGREES CENTIGRADE	•	18
VII	OXYGEN CONSUMPTION, CARBON DIOXIDE PRODUCTION AND RESPIRATORY QUOTIENTS OF INSTARS ONE THROUGH TEN AT TEN DEGREES CENTIGRADE	•	21
VIII	OXYGEN CONSUMPTION, CARBON DIOXIDE PRODUCTION AND RESPIRATORY QUOTIENTS OF INSTARS ONE THROUGH TEN AT FIFTEEN DEGREES CENTIGRADE	•	21
IX	OXYGEN CONSUMPTION, CARBON DIOXIDE PRODUCTION AND RESPIRATORY QUOTIENTS OF INSTARS ONE THROUGH TEN AT TWENTY DEGREES CENTIGRADE	•	22

ACKNOWLEDGMENTS

The author wishes to express sincere appreciation and thanks to Dr. John C. Wright for his encouragement, guidance and critical review throughout the research and writing of this thesis. Special thanks are also due Dr. John H. Rumely, Dr. H. S. MacWithey, Jr. and Dr. Louis D. S. Smith for their valuable suggestions and criticisms.

Thanks are due Dr. Rufus Kiser of Centralia Junior College for identifying the <u>Daphnia</u> used in this study.

Special thanks are due the National Science Foundation for their financial assistance and also the Department of Botany and Bacteriology for providing the necessary laboratory facilities. Thanks also to the capable faculty members of this department who helped to make this study possible.

Any errors or omissions in this thesis are the sole responsibility of the author.

ABSTRACT

A detailed study was made on the life history, natality, growth, and mortality of <u>Daphnia pulex</u> ssp. <u>pulicoides</u> Woltereck 1932. In addition, a grazing study was carried out at temperatures of 5° , 10° , 15° , 20° , and 25° C. and at instar levels one through ten. Grazing data is presented in tabular form and summarized with a graph. Temperature effect on grazing rates was noted. Respiration studies were carried out at temperatures of 10° , 15° , and 20° C. at instar levels one through ten. A Q_{10} was calculated for oxygen consumption and also for carbon dioxide production. The Q_{10} was between the temperature levels of 10° and 20° C. A disucssion and a review of literature is presented. Part V includes a short summary.

PART I

INTRODUCTION

The purpose of this study was to obtain data on the biology of Daphnia pulex ssp. pulicoides Woltereck, 1932 which could be applied to the calculation of secondary productivity in Canyon Ferry Reservoir. This subspecies is listed as Daphnia schodleri Sars, 1862 in Brook's monograph (1958) and is the major zooplankter in Canyon Ferry Reservoir, an artificial impoundment located on the Missouri River near Helena, Montana (Wright, 1958).

Although it was not the primary purpose of this study to calculate secondary productivity, certain factors concerning the biology of the secondary producer must be known before an accurate measurement of secondary productivity can be obtained. These factors include rates of growth, reproduction, mortality, grazing, and respiration. In order to arrive at a better knowledge of these factors, three studies were carried out -- a life history study, a grazing study and respiration studies (oxygen consumption and carbon dioxide production).

Financial assistance was obtained from National Science Foundation Research Grant No. 3063. Laboratory facilities were made available by the Botany and Bacteriology Department, Montana State College, Bozeman, Montana.

METHODS

Life History Study

Animals collected from Canyon Ferry Reservoir were brought to the laboratory. One female was placed in a two liter flask containing filtered lake water and a generous supply of Ankistrodesmus for food. This female was watched closely for the releasing of young first-instar animals. When the young were released, they were immediately obtained, measured and placed in a 200 ml. flask containing filtered lake water and a generous amount of Ankistrodesmus cells to insure an abundant supply of food. Thirty-six flasks, each containing one first-instar D. pulex, were set up. A lighted controlled-temperature cabinet maintained at 16°C. was used to keep the experimental flasks at a constant temperature.

Daily, following the commencement of the experiment, each animal was picked out, anesthetized with six drops of chlorobutanol administered with a pipette and measured. After the beginning of the reproductive phase, the eggs or embryos carried by each mother were counted. When a mother released young they were removed from the experimental flask and counted. Cast carapaces, increases in lenght, and the number of young released were used as criteria for the determination of the passing of the animals from one instar to the next. Additional cells of Ankistrodesmus were added from time to time in order to keep the experimental animals in a well-fed condition.

To verify the reliability of the data obtained from the original 36 animals, an additional series of 12 animals was run after the first series had been completed. Thus, the life histories of 48 <u>D. pulex</u> were followed day by day from birth to death

Grazing Study

The experimental animals were conditioned in filtered pond water for 24 hours prior to the commencement of each experiment. Various numbers of animals of a given instar were placed in a flask containing 100 ml. of filtered pond water. Log phase <u>Chlamydomonas</u> cells which were grown in liquid Modified Bristol's Solution (Bold, 1949) were centrifuged from the culture media to remove any toxic material which may have been produced by the algae. These cells were resuspended in the pond water contained in the experimental flasks. An attempt was made to obtain data on grazing rates of instars one through ten at temperature levels of 5°, 10°, 15°, 20°, and 25° C.

The cell concentration does not influence the filtering rates of zooplankton to any great extent. It is for this reason that cell concentrations were not considered as important as long as they were above the level of 0.15 million cells per milliliter (Ryther, 1954).

One milliliter aliquots were withdrawn at the start and at the finish of each experiment and placed in a Sedgewick-Rafter Counting Chamber. Fifty fields were counted using a Whipple micrometer disc as the boundaries of the field. The differences in cell concentrations were then applied to Gauld's equations (1951);

$$\frac{C_t}{C_0} = e^{-nk} \tag{1}$$

where C_{o} is the initial cell concentration, C_{t} the cell concentration at time (t), n the number of hours, and k the exponential function, and (2)

F is the filtering rate and V the volume of water per animal.

Experiments were limited to one to two hours to reduce the error brought about by the algae settling out (Ryther, 1954). The animals were counted and measured at the termination of each experiment. They were then placed in a weighed crucible, oven-dried at 100° C. for 16 hours and tared.

Respiration Studies

Oxygen Consumption Determinations

F = Vk

Oxygen uptake was measured by means of the polarometric method of Petering and Daniels (1938). A Fischer Elecdropode was used. The dropping mercury electorde was calibrated for dissolved oxygen concentrations by measuring the difference between galvanometer deflections at -0.1 volt and -1.0 volt. Oxygen concentration in the sample was determined by the Winkler method. Several calibrations were made at various oxygen concentrations. Oxygen concentrations were plotted against the corresponding galvanometer deflection differences and a regression line fitted to the points.

It was sometimes necessary to add a supporting electrolyte to the water sample. Potassium chloride (0.1 N) was used as the electrolyte when needed.

The animals used in the experiments were conditioned in filtered pond water. Each group of animals to be used in a given experiment was kept for 24 hours in pond water held at a temperature corresponding to that at which the experiment would be run. Twenty-five animals of approximately the same instar were picked from the conditioning water and placed in 125 ml. steam sterilized glass-stoppered bottles containing fresh filtered pond water. A control bottle lacking animals was set up to correct for microorganism respiration. Observations of oxygen uptake and carbon dioxide production were obtained at the beginning and again after 24 hours, at which time the experiments were terminated.

The animals were recovered at the end of each experiment and their lengths were measured with a microscope containing an ocular micrometer disc. They were then placed in a weighed crucible, oven-dried for 16 hours at 100° C., and weighed. Experiments were run at temperatures of 20° , 15° , and 10° at instar levels one through ten.

Carbon Dioxide Production Determinations

Carbon dioxide addition to water during animals respiration was measured by a modification of the method employed by Verduin (1956a). A Beckman model GS pH meter was employed in order to give a higher degree of accuracy. The Beckman GS pH meter has an expanded scale of 1,000 units which encompasses a pH range of three pH units. The expanded scale is calibrated in terms of millivolts which need not be converted to the pH scale in this case. Since one milliliter of 0.010 N NaOH is equivalent to 10 micromoles of carbon dioxide, the number of

micromoles of ${\rm CO}_2$ equivalent to a one unit change on the expanded scale can be calculated.

For example, if the expanded scale reading changed from 470 to 500 MV due to animal respiration and the reading became 460 MV after the addition of one milliliter of 0.010 N NaOH, then 500 minus 460 or 40 units is equivalent to 10 micromoles $\rm CO_2$ per liter. Hence one unit equals $\frac{10 \text{ micromoles}}{40}$ or 0.25 micromoles $\rm CO_2$ per unit. Animal respiration caused a change of 30 units (500 - 470); therefore, 30 times 0.25 micromoles $\rm CO_2$ per unit equals 7.50 micromoles $\rm CO_2$ per length of time covered by the experiment.

PART III

RESULTS

Life History Study

Instar-Weight Relationship

Figure 1 presents the instar-weight relationship by instar. The weights were obtained by selecting animals of each instar group and placing a known number on a tared cover slip. These animals were then oven-dried at 100° C. for 16 hours, cooled in a desiccator and weighed.

The instar weight relationship obtained in this expierment was similar to that found by Richman (1958) with <u>Daphnia pulex</u> var. <u>pulicaria</u> Forbes and Edmondson (1955) with <u>Daphnia pulex</u> var. <u>tenebrosa</u> Sars.

Growth Per Instar

Figure 2 shows the increase in length in millimeters per instar.

The slope of the line is negative; that is, the younger animals showed a greater rate of growth per instar than did the older animals.

Duration of Each Instar

The average time in days for passage of one instar to the next is shown in Figure 3. Here again, the age of the animal influenced the rate of change of this process. Instars one through five passed through each succeeding instar rather rapidly. Passage from instar one to instar two took only 1.39 days, while passage from instar four to instar five

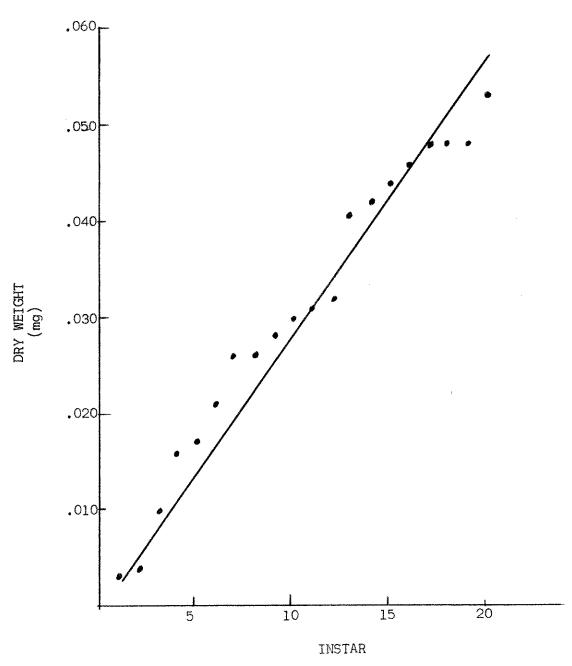


Figure 1. Instar-weight Relationship.

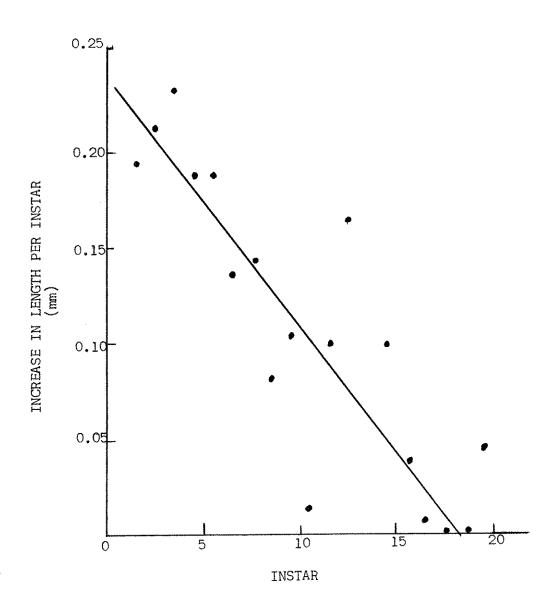


Figure 2. Average Increase in Length of Each Instar.

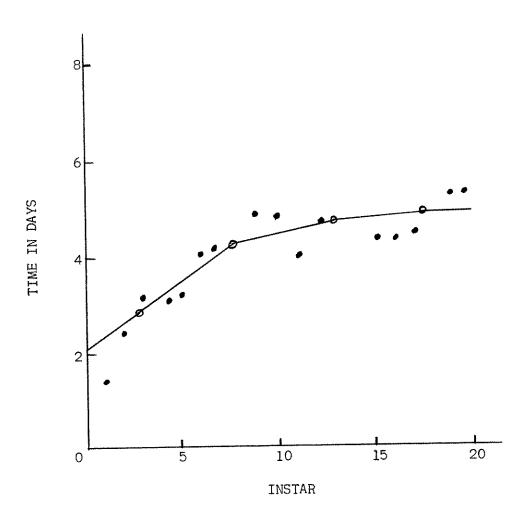


Figure 3. Average Time in Days Required for Passage from One Instar to the Next.

took 3.00 days. The duration of each instar increased until passage from instar 19 to instar 20 took 5.25 days.

Reproduction

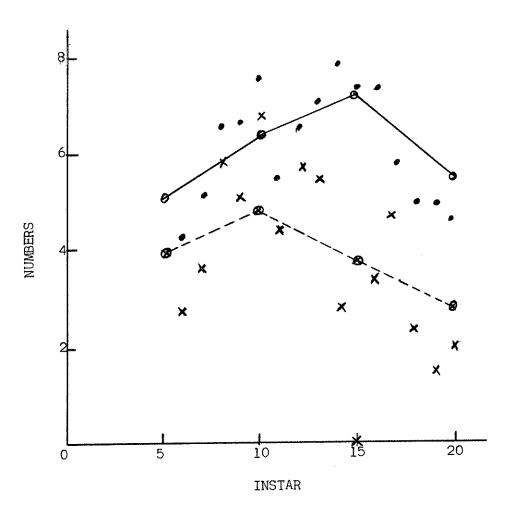
Reproduction generally began at the fith instar level and occasion—ally the sixth. When reproduction began, the duration of each instar was markedly increased over that of the younger, nonreproductive instars. In life history studies of <u>Daphnia longispina</u> (Wood and Banta, 1933 and Ingle, 1933) and <u>Daphnia magna</u> (Anderson and Jenkins, 1942), the experimental animals were also found to be primiparous at the fifth instar and less frequently at the sixth instar.

Figure 4 shows the average number of eggs formed and the average number of young released in each instar. Previous papers on life history studies of <u>Daphnia</u> have assumed that all eggs formed per instar were viable. This was not found to be the case with <u>Daphnia pulex</u> ssp. <u>pulicoides</u> in this study.

The highest percentage of viable eggs was 77 percent produced during the fifth instar. The lowest was 51 percent produced during the fifteenth instar. The over-all percentage of viability was 61.

Eggs that were formed but not viable took on a watery appearance, whereas viable eggs developed embryos. The nonviable eggs shrank in size and finally disappeared from the brood chamber by the time the young were to be released.

In some cases, the entire brood of eggs was attacked and destroyed by a fungal mycelium. This fungus did not appear to affect the mother as



Eggs Formed x--x Young Released

Figure 4. Average Number of Eggs Formed and Average Number of Young Released Per Reproductive Instar.

she would produce a brood comparable to other specimens during the following instar. The fungus was not studied in this investigation.

Mortality Rate

Figure 5 gives the number of survivors per instar. The highest mortality rates were encountered in the nonreproductive instars. Beginning with the reproductive instars, the curve tends to level off. The older the population became, the lower was the mortality rate of the population.

In Figure 5, data on 26 instars are included. Beyond the twentieth instar there were too few individuals to satisfactorily represent the population.

Table I gives pertinent data on instars one through 26. Seventy-six days elapsed between instars one and 20, and 73 days between instars 20 and 26, or a total of 149 days from birth to death.

Ingle (1933) using <u>Daphnia longispina</u> found 46.75 days to be the maximum longevity of his experimental animals. Anderson and Jenkins (1942) found <u>D. magna</u>, primiparous in the sixth instar, to have a life span of 53.54 days involving passage of 22 instars. Anderson and Zupancic (1933) reported <u>Daphnia pulex</u> as reaching the twentieth instar at which time the experiment was concluded. Ingle and Wood (1937) using <u>D. longispina</u> found the maximum longevity of their animals to be 51.19 days; this occurred among animals starved until the fifteenth instar. Their animals attained instar 23.

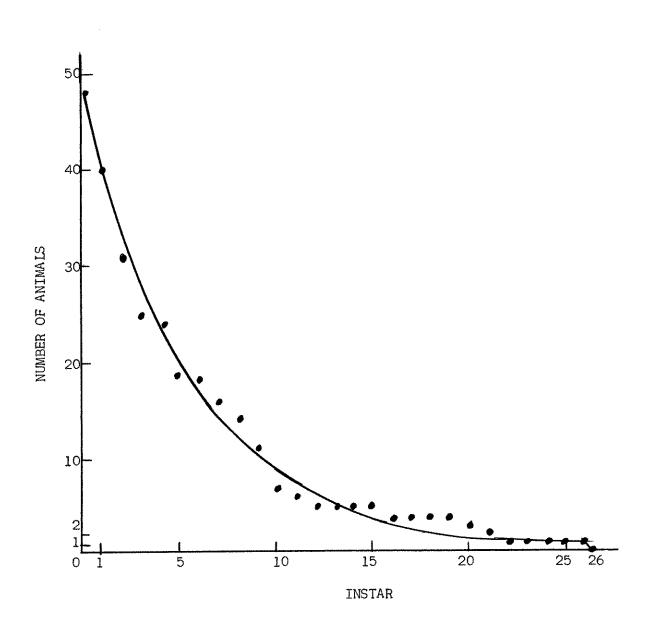


Figure 5. Survivors Per Instar.

TABLE I. DATA ON INSTARS ONE THROUGH TWENTY-SIX.

				Average			
		Average	Average	Dry	Average	Average	Average
	No.	Length	Increase	Weight	No.	No.	Time
Instar	Animals	(mm)	(mm)	(mq)	Eggs	Young	(days)
1	48	.6603		.003	-		1.39
2	40	.8579	.1976	.004	····	JOHNS KNASK WINAA	2.32
3	31	1.0703	.2124	.010			
J 4	25	1.3033	.2330	.016			3.10 3.00
5	24	1.4826	.1793	.017	5.08	3.92	
6	19	1.6615	.1789	.021	4.33	2.77	3.30
7	18	1.7977	.1362	.026	5.11	3.65	4.05
8	16	1.9414	.1437	.026	6.60	5.87	4.13
9	14	2.0247	.0833	.028	6.71	5.07	4.07
10	11	2.1287	.1040	.030	7.63	6.81	4.91
						4 40	4.70
11	8	2.1488	.0158	.031	5.50	4.40	4.00
12	6	2.2489	.1001	.032	6.60	5.75	
13	5	2.4142	.1653	.041	7.14	5.57	4.50
14	5	2.3889	.2053	.042	8.60	2.80	4.60
15	5	2.4901	.1012	.044	7.40	0.00	4.80
16	5	2.5280	.0379	.046	7.40	3.40	4.20
17	4	2.5359	.0079	.048	5.80	4.60	4.20
18	4	2,5359	.0000	.048	5.00	2.25	4.25
19	4	2.5359	.0000	.048	5.00	1.50	5.25
20	4	2.5833	.0474	.053	4.75	2.00	5.25
			0000	05.0	E 00	2.80	6.00
21	3	2.5833	.0000	.053	5.00		6.21
22	2	2.5916	.0083	.055	5.00	3.00	
23	1	2.6544	.0628	.058	8.00	6.00	9.00
24	1	2.7492	.0948	.060	10.00	6.00	10.00
25	1	2.8440	.0948	.065	6.00	6.00	14.00
26	1	2.9072	.0628	.068	6.00	6.00	27.00

Grazing Study

Tables II, III, IV, V and VI give the results of the experiments conducted on grazing rates for each instar at each temperature level.

TABLE II. VOLUME OF WATER FILTERED BY <u>DAPHNIA</u> INSTARS ONE THROUGH TEN AT FIVE DEGREES CENTIGRADE.

Instar	Length (mm)	Dry Weight (mg)	Volume Filtered (ml/Daphnia/ day)	Volume Filtered (ml/mg/day)
<u>, , , , , , , , , , , , , , , , , , , </u>	-70	.0050	6.72	1,344
1	.70		5.54	615
2	.87	.0090	7.76	715
3	1.09	.0114	8.54	635
4	1.27	.0137		578
5	1.49	.0160	9.24	6 3 9
6	1.68	.0170	10.86	
7	1.77	.0180	12.65	703
8	1.86	.0187	13.00	727
9	2.01	.0210	10.38	494
1Ó	2.14	.0236	10.49	455
lverage	1.49	.0153	9.52	691

TABLE III. VOLUME OF WATER FILTERED BY <u>DAPHNIA</u> INSTARS ONE THROUGH TEN AT TEN DEGREES CENTIGRADE.

Instar	Length (mm)	Dry Weight (mg)	Volume Filtered (ml/Daphnia/ day)	Volume Filtered (ml/mg/day)
		0005	9.12	2,606
1	.74	.0035		2,932
2	.94	.0050	14.66	•
3	1.12	.0105	11.56	1,120
4	1.33	.0157	10.66	707
5	1.48	.0183	16.47	884
6	1.68	.0190	13.18	694
7	1.78	.0210	11.71	564
	1.92	.0256	17.39	679
8 9	1.99	.0270	19.12	697
		.0273	16.99	670
10	2.14	•0210	10.77	
Average	1.51	.0173	14.09	1,155

TABLE IV. VOLUME OF WATER FILTERED BY DAPHNIA INSTARS ON THROUGH TEN AT FIFTEEN DEGREES CENTIGRADE.

Instar	Length (mm)	Dry Weight (mg)	Volume Filtered (ml/Daphnia/ day)	Volume Filtered (ml/mg/day)
				2 452
1	.68	.0040	11.67	3,453
2	.86	.0090	21.00	2,333
3	1.03	.0110	18.60	1,690
4	1.30	.0143	14,00	951
5	1.54	.0180	21.60	1,200
6	1.66	.0210	20.40	971
7	1.74	.0280	18.00	643
	1.86	.0300	18.96	699
8	1.93	0320	32.40	1,013
9		.0360	28.80	800
10	2.15	.0300	~~ • O O	
Average	1.48	.0203	20.54	1,375

TABLE V. VOLUME OF WATER FILTERED BY <u>DAPHNIA</u> INSTARS ONE THROUGH TEN AT TWENTY DEGREES CENTIGRADE.

Instar	Length (mm)	Dry Weight (mg)	Volume Filtered (ml/Daphnia/ day)	Volume Filtered (ml/mg/day)
			17.07	2,570
1	.71	.0070	16.86	•
2	.87	.0090	16.86	1,874
3	1.10	.0125	17.28	1,384
4	1.33	.0165	19.20	1,154
5	1.50	.0205	22.81	1,110
6	1.67	.0260	22,20	85 0
7	1.81	.0300	26.98	896
	1.97	.0330	23.26	705
8		.0340	30.00	882
9	2.00	_	29.15	776
10	2.23	.0375	27.10	, , ,
Average	1.52	.0228	22.46	1,220

TABLE VI. VOLUME OF WATER FILTERED BY <u>DAPHNIA</u> INSTARS ONE THROUGH TEN AT TWENTY-FIVE DEGREES CENTIGRADE.

Instar	Length (mm)	Dry Weight (mg)	Volume Filtered (ml/Daphnia/ day)	Volume Filtered (ml/mg/day)
•	.70	.0055	10.81	1,954
1 2	.88	.0060	15.10	2,517
3	1.08	.0115	17.06	1,493
4	1,33	.0165	17.77	1,079
5	1.48	.0180	21.17	1,176
6	1.62	.0182	25.96	1,455
7	1.79	.0215	24.98	1,342
8	1.92	.0260	30.00	1,154
9	2.04	.0300	24.72	824
10	2.16	.0320	28.08	887
Average	1.50	.0175	21.57	1,388

It was apparent that temperature conditioning is a definite factor in the filtering rates of <u>Daphnia pulex</u> at all instar levels.

The maximum filtering rates on the basis of ml/Daphnia/day were always encountered at either the eighth or ninth instar. The maximum filtering rate per animal was encountered during the eight instar at 5° and 25° C. while the maximum filtering rate at 10°, 15° and 20° C. occurred at the ninth instar level. Ryther (1954) found a similar relationship with Daphnia magna since the filtering rate per animal increased up to animals of 0.12 mg. dry weight in size. Animals of 0.13 to 0.15 mg. dry weight had lower filtering rates per animal than did the smaller animals.

The filtering rate on a ml/mg dry weight/day basis was an inverse function of size, the smaller animals having a higher filtering rate+than did the larger animals. This is in close agreement with all studies previously mentioned.

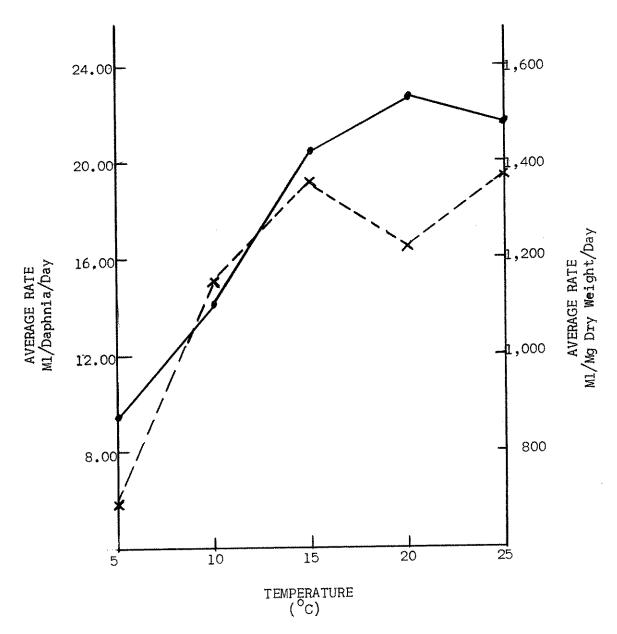
Figure 6 is the synopsis of all experiments. Each point represents the average values obtained from each temperature level. It can be noted that there is a maximum in the filtering rate per milligram dry weight at 15° C. and a decrease at 20° C. Conover (1956) found a relationship of this type with <u>Acartia clausi</u> and <u>A. tonsa</u>.

Respiration Studies

In general, oxygen consumption and carbon dioxide production increased with increase in body size, but decreased per animal on a unit weight basis (Tables VII-IX). However, there was considerable scatter in the data on respiration for large numbers of samples had to be obtained in order to get reliable averages.

Figure 7 gives the respiration rate by instar per milligram dry weight per hour. The smaller animals had a higher respiration rate than did the larger animals. The same relationship was found to hold true for <u>D. pulex ssp. pulicaria</u> (Richman, 1958) and <u>Acartia clausi</u> and <u>A. tonsa</u> (Conover, 1956).

Oxygen consumption varied from 5.290 micromoles/mg dry weight/hour for first instar animals to 0.561 micromoles/mg dry weight/hour for tenth instar animals. Carbon dioxide production varied from 5.475 micromoles/mg dry weight/hour for first instar animals to 0.511 micromoles/mg dry weight/hour for tenth instar animals.



.____. Ml/Daphnia/Day x---x Ml/Mg Dry Weight/Day

Figure 6. Average Filtering Rates of <u>Daphnia pulex</u> at Each Temperature Level.

TABLE VII. OXYGEN CONSUMPTION, CARBON DIOXIDE PRODUCTION AND RESPIRATORY QUOTIENTS OF INSTARS ONE THROUGH TEN AT TEN DEGREES CENTIGRADE.

tr () transfere til () e transfere til () e tr	Dry			ygen umption	Carbon Produ	over the second	
Instar	Length (mm)	Wt. (mg)	uM/A/hr	µM/mg dry wt/hr	μM/A/hr	μM/mg dry wt/hr	RQ
1	.7110	.0030	.0076	2,530	.0127	4.230	1.67
2	.8658	.0060	.0098	1.630	.0122	2.030	1.25
3	1.0618	.0130	.0101	0.780	.0123	0.950	1.22
4	1.2510	.0150	.0310	2.067	.0260	1.733	0.84
5	1.4347	.0210	.0245	1.296	.0231	1.190	0.92
6	1.6084	.0230	.0095	0.410	.0114	0.500	1.22
7	1.8202	.0300	.0105	0.350	.0133	0.440	1.26
8	1.9120	.0320	.0108	0.338	.0138	0.431	1.28
9	2.0256	.0310	.0109	0.350	.0139	0.450	1.29
10	2.1867	.0320	.0114	0.360	.0126	0.390	1.08
Average	1.4777	.0206	.0136	0.911	.0151	1.234	1.20

TABLE VIII. OXYGEN CONSUMPTION, CARBON DIOXIDE PRODUCTION AND RESPIRATORY QUOTIENTS OF INSTARS ONE THROUGH TEN AT FIFTEEN DEGREES CENTIGRADE.

***************************************		Dry	Oxygen Consumption			Dioxide uction	
·	Length	Wt.	4.31/5/5	μM/mg dry		M/mg dry	DΩ
Instar	(mm)	(mg)	MM/A/hr	wt/hr	#M/A/hr	wt/hr	RQ
1	.7079	.0034	.0190	5.483	.0210	6.053	1.09
2	.8658	.0046	.0140	3.091	.0170	3.696	1.20
3	1.0365	.0111	.0305	3.146	.0270	2.593	1.15
4	1.2976	.0172	.0150	0.893	.0240	1.395	1.56
5	1.4220	.0167	.0363	2.263	.0363	2.263	1.00
6	1.6379	.0244	.0147	0.605	.0147	0.605	1.00
7	1.8044	.0300	.0145	0.477	.0140	0.467	0.98
8	1.9608	.0316	.0155	0.489	.0150	0.475	0.98
9	2.0461	.0340	.0155	0.461	.0160	0.476	1.02
10	2.1614	.0352	.0170	0.478	.0160	0.455	0.95
Average	1.4940	.0208	.0192	1.739	.0201	1.847	1.09

TABLE IX. OXYGEN CONSUMPTION, CARBON DIOXIDE PRODUCTION AND RESPIRATORY QUOTIENTS OF INSTARS ONE THROUGH TEN AT TWENTY DEGREES CENTIGRADE.

		Dry	Oxygen Consumption		Carbon Dioxide Production		
Instar	Length (mm)	Wt. (mg)	μM/A/hr	μM/mg dry wt/hr	μM/A/hr.	&M/mg dry wt/hr	RQ
1	.7126	.0035	.0275	7.857	.0215	6.143	0.79
2	.9496	.0069	.0295	4.542	.0300	4.500	1.01
3	1.1660	.0150	.0380	2.533	.0360	2.250	0.89
4	1.3430	.0160	.0220	1.375	.0210	1.313	0.95
5	1,4220	.0160	.0230	1.438	.0220	1.375	0.96
6	1.6795	.0205	.0325	1.673	.0310	1.621	0.97
7	1.7633	.0250	.0270	1.080	.0240	0.960	0.89
8	1.9118	.0287	.0250	0.871	.0203	0.713	0.82
9	2.0350	.0291	.0260	0.893	.0210	0.723	0.81
10	2.1477	.0320	.0270	0.844	.0220	0.688	0.82
Average	1.5131	.0193	.0278	2.311	.0249	2.129	0.90

The average respiration rates of all instars at each temperature level on a per animal basis are represented in Figure 8. Temperature appeared to be a definite factor in the respiration of \underline{D} . \underline{pulex} .

The average respiration rates of all instars on a milligram dry weight basis are shown in Figure 9. At 10° C. the average rate for oxygen uptake was 0.911 micromoles/mg dry weight/hour for animals having an average weight of 0.0206 milligram. Carbon dioxide production for animals of the same average weight and at 10° C. was 1.234 micromoles/mg dry weight/hour. The respiration rate increased to 2.311 micromoles/mg dry weight/hour for oxygen uptake at 20° C. and 2.129 micromoles/mg dry weight/hour for carbon dioxide production at 20° C. The average dry weight per animal for the 20° C. series was 0.0193 milligram. The Q_{10}

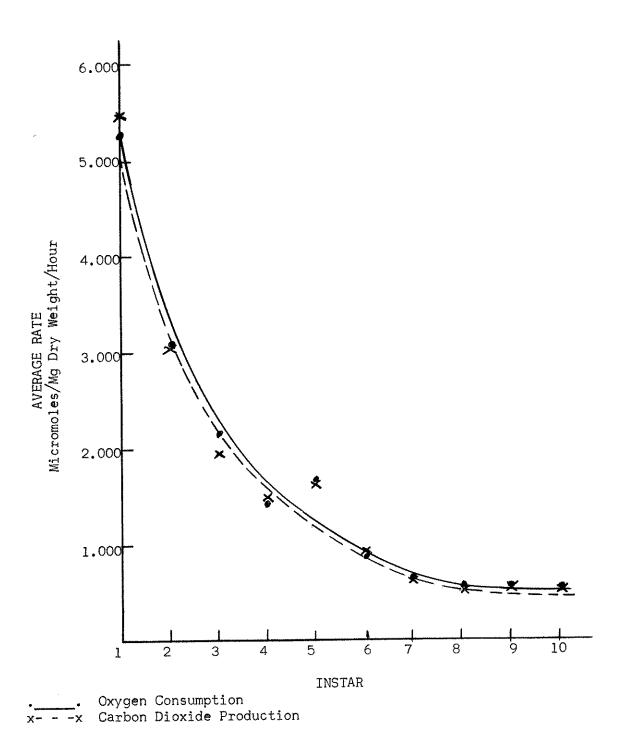
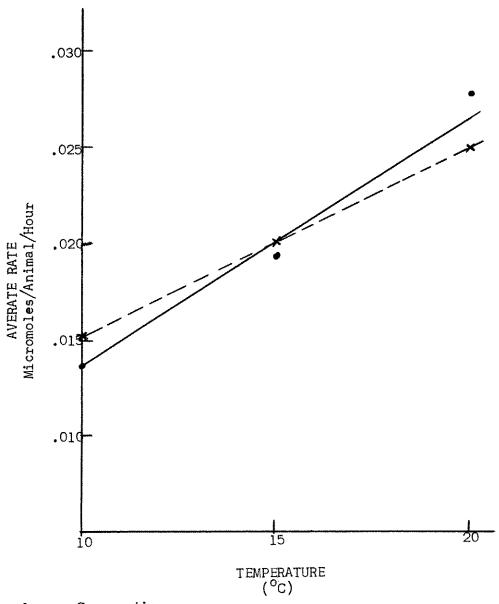
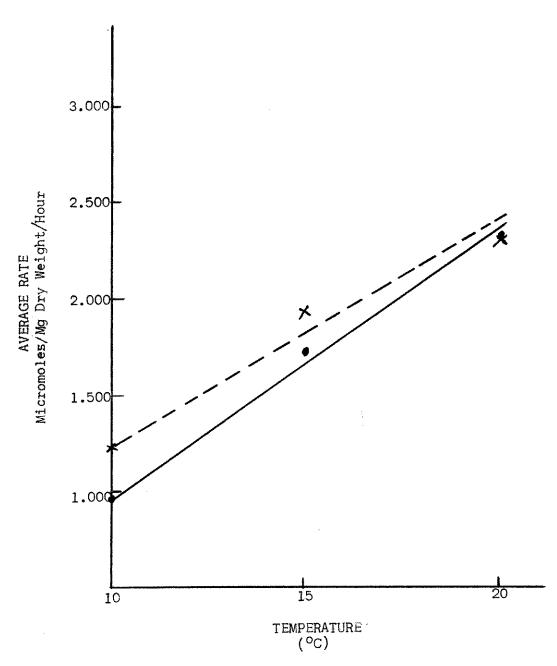


Figure 7. Average Rates of Oxygen Consumption and Carbon Dioxide Production -- by Instar.



.___. Oxygen Consumption x--x Carbon Dioxide Production

Figure 8. Average Rate of Oxygen Consumption and Carbon Dioxide Production on a Per Animal Basis at Each Temperature Level.



• Oxygen Consumption
-x- -x Carbon Dioxide Production

Figure 9. Average Rate of Oxygen Consumption and Carbon Dioxide Production per Unit Weight at Each Temperature Level.

for oxygen consumption was 2.53 and that for carbon dioxide production was 1.73. Acartia was found to have a \mathbf{Q}_{10} of 2.04 by Conover (1956) between temperatures of 10° and 20° C.

Oxygen consumption and carbon dioxide production corresponded closely at all temperature levels. The increase in both rates was essentially linear. The mean oxygen consumption rate was found to be 0.0202 micromoles/animal/hour and 1.654 micromoles/mg dry weight/hour while the mean carbon dioxide production rate was 0.0200 micromoles/animal/hour and 1.737 micromoles/mg dry weight/hour. The overall respiratory quotient was 1.031 which corresponds closely to 1.03 found for <u>Daphnia pulex</u> ssp. pulicaria (Richman, 1958).

PART IV

DISCUSSION

Life History Study

Life history studies have been carried on with various species of Daphnia (Anderson and Jenkins, 1942; Anderson and Zupancic, 1933; Ingle, 1933; Ingle, Wood and Banta, 1937; and Wood and Banta, 1936). The data collected in this study were essentially the same as were found in the studies previously mentioned except for three previously unreported points. These are (1) production of nonviable eggs, (2) fungus mycelium in the brood chamber and (3) a longevity of 149 days.

In previous studies the number of young released was assumed to be the same as the number of eggs formed for that brood or the number of eggs formed per brood was taken as the number of young that would be produced. In the present study it was discovered that some eggs which were not viable were produced. These eggs disappeared before the young were released.

The fungus that attacked brood chambers completely destroyed all eggs formed for that instar. The fungus did not affect the succeeding instar. In no case did the fungus affect the development taking place in the brood chamber after the eggs had passed into the embryonic stages. When the fungus was present, the mycelium replaced the eggs in the brood chamber.

The life span of 26 instars persisting through 149 days is the longest reported for <u>Daphnia</u>. This span was represented by only one animal of the

original 48. This one animal is probably exceptional because of the fact that it lived 60 days longer than any of the other original animals. It died after remaining in the twenty-sixth instar for 27 days.

Grazing Study

The overall average rate of all instars at all temperature levels was 1.166 L/mg dry weight/day. This is similar to Wright's (1958) grazing coefficient of 1.064 L/mg dry weight zooplankton/day and his calculation from Ryther's data of 1.060 L/mg dry weight zooplankton/day. Conover (1956) found rates of Acartia clausi and A. tonsa in the range of 1.20 to 1.40 L/mg dry weight/day which is in close agreement with the 1.166 L/mg dry weight/day found for <u>Daphnia</u> <u>pulex</u>. Richman (1958) using <u>Daphnia</u> pulex ssp. pulicaria as the experimental animal and palmella Chlamydomonas found filtering rates from 176-330 ml/mg dry weight/day. These values were obtained at 20° C. for 24 hours. It is evident that Richman's rates are noticably lower than those found for Daphnia pulex ssp. pulicoides at any temperature. When \underline{D} . \underline{pulex} ssp. $\underline{pulicoides}$ were subjected to exactly the same conditions as Richman used, similar rates (100-381 ml/mg dry weight/ day) were found for animals of similar size. Ryther pointed out in his paper that algae tend to settle out if the grazing experiments were allowed to run over one to two hours. Daphnia is not a scavenger and generally does not search for food which has settled out of the water.

Palmella <u>Chlamydomonas</u> forms macroscopic colonies that resist fragmentation even under vigorous agitation. The question which occurs is whether or not the palmella colonies are of such a size that they may be efficiently taken into the intestinal tract of <u>Daphnia</u>. Ryther investigated the relationship between particle size and filtering rate and found that cell volumes of $3.5-25.5\mu^3$ were filtered at equal rates.

Respiration Studies

Considerable differences in oxygen consumption rates for both marine and freshwater zooplankton have been found. The rate of oxygen consumption ranges from 0.72 microliters/mg dry weight/hour with <u>D. magna</u> (MacArthur and Baille, 1929) up to 14.20 microliters/mg dry weight/hour for <u>Simocephalus expinosus</u> (Obreshkove, 1930).

Richman (1958) found an oxygen consumption rate for <u>Daphnia pulex</u> ssp. <u>pulicaria</u> of 7.21 microliters/mg/hour for animals over 1.0 mm. in length at 20° C. This compares with 19.264 micorliters/mg/hour found with <u>D. pulex</u> ssp. <u>pulicoides</u> of comparable size in the 20° C. series. Conover (1956) using <u>Acartia tonsa</u> and <u>A. clausi</u> found oxygen consumption rates of 8.12 to 12.06 microliters/mg/hour for his animals at 20° C.

The oxygen consumption rate found for <u>D</u>. <u>pulex</u> ssp. <u>pulicoides</u> at 10° C. was approximately 0.500 micromoles/mg/hour or 11.20 microliters/mg/hour for animals over 1.0 mm. in length. This figure was of the same magnitude as the previously mentioned rates. The rates of oxygen consumption for animals of 1.0 mm. or greater in length at 15° C. was 16.80 microliters/mg/hour. The mean of all rates is 37.04 microliters/mg/hour.

Carbon dioxide production was proportional to oxygen consumption.

Tenth instar animals produced 0.511 micromoles/mg dry weight/hour of

carbon dioxide with an oxygen uptake of 0.561 micromoles/mg/hour. Wright (1958) found a rate of 0.636 micromoles/mg/hour for <u>Daphnia pulex</u>. His animals had an average weight of 0.037 milligrams.

LITERATURE CITED

- Anderson, B. G., and J. C. Jenkins. 1942. A time study of the events in the life span of <u>Daphnia magna</u>. Biol. Bull. 83:260-272.
- Anderson, B. G., and L. J. Zupancic. 1933. Growth in <u>Daphnia pulex</u>.

 Anatom. Rec. 57 (4):27-50.
- Bold, H. C. 1949. The morphology of <u>Chlamydomonas Chlamydogama</u> sp. nov. Bull. Torrey Bot. Club 76:101-108.
- Brooks, J. L. 1957. The systematics of North American <u>Daphnia</u>. Conn. Acad. Arts and Sciences. 13:68-75.
- Conover, R. J. 1956. Oceanography of Long Island Sound, 1952-1954. VI Biology of <u>Acartia clausi</u> and <u>A. tonsa</u>. Bull. Bingham Oceanogr. Coll. 15:156-233.
- Edmondson, W. T. 1955. The seasonal life history of <u>Daphnia</u> in an Arctic lake. Ecology 36 (3):439-455.
- Gauld, D. T. 1951. The grazing rate of planktonic copepods. J. Mar. Biol. Ass. U.K., 29:695-706.
- Ingle, L., T. R. Wood, and A. M. Banta. 1937. A study of longevity, growth, reproduction and heart rate in <u>Daphnia longispina</u> as influenced by limitations in quantity of food. Jour. Exp. Zool. 76:325.
- MacArthur, J. W. and W. H. T. Baille. 1929. Metabolic activity and duration of life. II Metabolic rates and their relation to longevity in <u>Daphnia magna</u>. Jour. Exp. Zool. 53:221-242.
- Obreshkove, V., 1939. Oxygen consumption in the developmental stages of a Cladoceran. Physiol. Zool. 3:271-282.
- Petering, H. G., and F. Daniels. 1938. The determination of dissolved oxygen by means of the dropping mercury electrode, with applications in biology. J. Amer. Chem. Soc. 60:2796-2802.
- Richman, Sumner. 1958. The transformation of energy by <u>Daphnia pulex</u>. Ecol. Monogr. 28 (3):273-291.
- Ryther, J. H. 1954. Inhibitory effects upon the feeding of <u>Daphnia magna</u> with reference to growth, reproduction, and survival. Ecology. 35:522-533.

- Verduin, J. 1956. Energy fixation and utilization by natural communities in western Lake Erie. Ecology. 37:40-50.
- Wood, T. R., and A. M. Banta. 1936. Interpretations on data on growth and reproduction (<u>Daphnia longispina</u>). (abstract). Anat. Rec. 67:55.
- Wright, J. C. 1958. The limnology of Canyon Ferry Reservoir. Limnol. and Oceanogr. 3 (2):150-159.